

0040-4020(94)00768-3

Synthesis of 2'-Deoxy-Pyranosyl Nucleosides from Glycals through Consecutive Addition of Phenylselenenyl Chloride and Glycosylation. A Study of Factors Controlling the Stereoselectivity

Anas El-Laghdach, M^a Isabel Matheu, Sergio Castillón^{*}

Departament de Química, Universitat Rovira i Virgili, Pça. Imperial Tarraco 1, 43005 Tarragona, Spain

Abstract: 2'-Deoxy-2'-phenylselenenyl-pyranosyl nucleosides have been synthesised in a stereoselective way starting from glycals using selenium reagents, and converted into 2'-deoxynucleosides by treatment with tributyltin hydride. The stereoselectivity of the reaction has been shown to be dependent on the protecting groups, the structure of the starting glycal, the phenylselenenyl reagent and the solvent. Nucleosides of β -gluco β -galacto and α -arabino configuration are principally obtained, starting from the corresponding benzyl protected glycals, using PhSeCl as an activating reagent and ether as the solvent.

Nucleoside analogues are among the more active antiviral drugs.¹ The increasing importance of viral diseases has prompted a growing interest in nucleoside chemistry. In this way, pyranosyl nucleosides have been the object of attention as analogues of furanosyl nucleosides of proved antiviral activity,² as precursors in the synthesis of acyclo-nucleosides³ and in the preparation of oligopyranosyl nucleotides.⁴ Most of these reports deal with the synthesis of 2'-deoxy-&-pyranosyl nucleosides; however, there are no general solutions for the stereoselective synthesis of these compounds based on the glycosylation reaction, although the ß isomer is thermodynamically more stable.

2'-Deoxynucleosides are usually prepared by deoxygenating the 2'-position by way of a Barton type reaction starting from appropriate protected nucleosides;⁵ however, this method needs long protecting sequences to be applied for pyranosyl nucleosides.^{3a} Good results have been achieved starting from 1- α -bromo-2-deoxy-3,4,6-tri-(*p*-nitrobenzoyl)-D-arabino-hexo-pyranose,⁶ but S_N2 like pathways must be strictly guaranteed and this methodology has hardly been used in pyrimidine like nucleoside synthesis. During the seventies, extensive studies were developed oriented towards the synthesis of 2',3'-dideoxi-2',3'-eno-pyranosyl nucleosides from glycals, some of which had shown antibiotic properties, through a S_N2' type reaction catalysed by Lewis acids.⁷ Recently, 2',3'-dideoxi-2',3'-eno nucleosides have been obtained from 2',3'-dideoxi-2',3'-eno glycosides by a condensation reaction catalysed by Pd(0) catalyst.⁸

In the glycoside field, good stereoselectivity in the synthesis of 2'-deoxyglycosides has been obtained via sulphur⁹ selenium¹⁰ and iodine¹¹⁻¹³ mediated glycosylation reactions starting from glycals. In this context, and

given the strong stereoselectivity dependence of this reaction from the starting glycal, solvent, nucleophile, etc., we decided to undertake a systematic study looking for the best reaction conditions in the stereoselective synthesis of 2'-deoxy-8-pyranosyl nucleosides starting from glycals and using selenium reagents.

2'-DEOXY-2'-PHENYLSELENENYL-PYRANOSYL NUCLEOSIDES

Preliminary experiments carried out by the reaction of tri-O-acetyl-D-glucal (1a) with PhSeCl and bis-(trimethylsilyl)uracil in different solvents (methylene chloride, acetonitrile, ether) and under different reaction conditions gave only PhSeCl addition products; PhSeBr and PhSeI showed similar results. The reason for the different behaviour between alcohols and bis(trimethylsilyl)uracil must be explained by the lower nucleophilicity of this last reagent in comparison with the alcoholate.

Increasing reagent concentrations or employing N-phenylselenenylphthalimide (NPSP), a selenium reagent with a less nucleophilic counterion, also gave negative results. The use of silver triflate as a halogen captor, allowed us to obtain 2'-deoxy-2'-phenylselenenyl nucleosides¹⁴ 4a and 5a together with the 2',3'-dideoxy-2',3'-eno derivative 6^{7e} (Table 1, Entry 1). Compounds 4a and 5a, both with a *trans* disposition of uracil and the phenylselenenyl group, were obtained by the attack of the phenylselenenyl reagent on both faces of the alkene and subsequent opening of the phenylselenonium cations formed (Scheme 1). Working at room temperature (Table 1, Entry 2) or using other solvents (Table 1, Entry 3), neither the yield nor the selectivity were improved. The best yields were obtained by increasing the glycal/PhSeCl/AgOTf ratio and avoiding the aqueous work-up (Table 1, Entry 4), although the selectivity was always low.

Compound 6, was probably obtained by a S_N2' type reaction⁷ (Ferrier rearrangement) catalysed by the TMSOTf formed in the reaction medium.

		N	lolar Rat	io					
Entry	Glucal	PhSeCl	AgOTf	Uracil(TMS)2	Solvent	Temp.	Time	Yielda	4a/5a/6 ^b
1	1	1.2	1	2	CH ₂ Cl ₂	-200°C	3	50	45/45/10
2	1	1.2	1	2	CH ₂ Cl ₂	rt	1.2	47	45/45/10
3	1	1.2	1.2	2	CH ₃ CN	-200°C	22	35	45/45/10
4	1	1.5	1.7	2	Ether	rt	1	80	55/30/15

Table 1. Reaction of bis-(trimethylsilyl)uracil with 3,4,6-tri-O-acetyl-D-glucal (1a) induced by PhSeCl and silver triflate.

^a Expressed as a percentage of recovered mixture of products after chromatography. ^b Determined by H-1' integration in the ¹H NMR of the reaction mixture

The formation of elimination products was prevented by utilising ether type protecting groups. Thus, starting from tri-O-benzyl-D-glucal (1b) and carrying out the reaction in the best conditions as before, (Table 2, Entry 1), compounds 4b and 5b were obtained in good yields as an inseparable mixture.



Table 2. Reaction of bis-(trimethylsilyl)uracil with 3,4,6-tri-O-benzyl-D-glucal (1b) induced by PhSeCl and AgOTf.^a

		M	lolar Ratic						
Entry	Glucal PhSeCl		Catalyst Uracil(TMS) ₂		Solvent	Temp.	Time(h)	Yield(%) ^b 4b/5b(%	
1	1	1.5	1.7 ^d	2	CH ₂ Cl ₂	-40°Crt	3.5	87	50/50
2	1	1.5	1.7d	2	C ₆ H ₆	rt	1	88	80/20
3	1	1.5	1.7 ^d	2	CH ₃ CN	0°Crt	15	70	40/60
4	1	1.5	1.7 ^d	2	ether	0°Crt	0.4	85	82/18
5	1	1.5	1.7e	2	CH ₂ Cl ₂	-40°Crt	3.5	50	50/50
6	1	1.5	2.3 ^f	2	ether	0°Crt	24	60	84/16
7	1	1.5	4.6 ^f	2	benzene	rt	12	76	82/18

^a 0.25 mmolar solutions of sugar were used. ^b Expressed as a percentage of the recovered mixture of diastereomers after chromatographic separation. ^c Determined by integration of the H-1' protons in the ¹H NMR of the reaction mixture. ^dAgOTf. ^eSbC15. ^fTMSOTf.

In order to improve the stereoselectivity we undertook a systematic study of the reaction conditions. When glucal 1b was treated with uracil(TMS)₂ and PhSeCl in dichloromethane, at room temperature as well as at -60°C, no variations in stereoselectivity were observed. However, the stereoselectivity was shown to be



Table 3. Selenium reagent influence in the reaction of bis-(trimethylsilyl)uracil with 3,4,6-tri-O-benzyl-D-glucal (1b).^a

Entry	PhSeX	Solvent	Time(h)	Yield(%)	4b/5b(%) ^t
1	PhSeC1 ^c	Ether	0.4	85	82/18
2	PhSeBr ^c	Ether	0.5	85	60/40
3	PhSeId	Ether	1	80	50/50

^a 0.25 mmolar solutions of sugar were used. ^b Determined by integration of the H-1' protons in the ¹H NMR of the reaction mixture. ^c Molar ratio sugar/PhSeX/AgOTf/Uracil was 1/1.5/1.7/2. ^d Molar ratio sugar/PhSeI/AgOTf/Uracil was 1/2/2.

Starting Glycal ^a	Solvent	Yield(%) ^b	2'-Phenyiselenenyi	nucleosides (%) ^c
AcO			ß-gluco	o-manno
	Ether	68	4a (66)	5a (34)
OAc	Acetonitrile	32	4a (50)	Sa (50)
BnO.	Ether	85	4h (80)	Sh (20)
	Acetonitrile	70	4b (40)	5b (60)
AcO			ß-galacio	a-talo
	Ether	80	7a (50)	8a (50)
	Acetonitrile	40	7a (25)	8a (75)
	Ether	82	7b (91)	8b (9)
OBn (20)	Acetonitrile	37	7b (83)	8b (17)
4-0			oi-arabino	ß-ribo
	Ether	75	10a (60)	9a (40)
Aco (34)	Acetonitrile	65	10a (53)	9a (47)
	Ether	71	1 0b (92)	9b (8)
BnO (30)	Acetonitrile	42	10b (50)	9b (50)

Table 4. Stereoselectivity in the synthesis of 2'-deoxy-2'-phenylselenenyl-pyranosyl nucleosides.

^a Shown in the preferred conformation. ^b Referred to the mixture of phenylselenenyl nucleosides. ^c Determined by integration of H-1' protons in the ¹H NMR spectra of the reaction mixture

dependent on the solvent. Thus, the β isomer was principally obtained when the reaction was carried out in ether or benzene, while the α isomer was the main one obtained in acetonitrile (Table 2, Entries 2, 3 and 4). Other chlorine activators such as SbCl₅ or TMSOTf (Table 2, Entries 5, 6 and 7), gave generally lower yields and slower reaction rates, but had no influence in the stereoselectivity.

The observed stereoselectivity must be related to the ratio of 1-halo-2-phenylselenenyl-glucopyranosyl derivatives present in the solution before the addition of silver triflate; therefore, the type of phenylselenenyl reagent should influence, because of steric or stereoelectronic factors, the equilibrium between the 2'-phenyl-selenenyl halopyranoses (Scheme 1). A study performed with PhSeCl, PhSeBr and PhSeI showed (Table 3) that the phenylselenenyl reagent had no influence on the yield, but it had a drastic influence on the stereoselectivity. The best results were obtained when PhSeCl was used.

Treatment of 3,4,6-tri-O-acetyl-D-galactal (2a) with PhSeCl, AgOTf and bis-(trimethylsilyl)uracil in CH₂Cl₂ resulted in to a 1:1 mixture of 2'-deoxy-2'-phenylselenenyl nucleosides 7a and 8a in 70% yield. The reaction in ether was much faster than in CH₂Cl₂, although a similar mixture of compounds was also obtained in 80% yield. The use of acetonitrile gave preferentially the product with α -manno configuration, resulting from the selenium attack above the molecular plane (8 face) (Scheme 1), 1:3 being the 7a/8a ratio. In this case the product resulting from the Ferrier rearrangement was not observed. Curiously, the selectivity for the tri-O-

J/Compound	4a	4b	5a	5 b	7a	7 b	8a	8b	9a	9b	10 a	10b
J _{1',2'}	11.0	10.7	10.8	10.9	10.7	10.6	10.7	9.0	10.7	1.9	10.7	10.5
J _{2',3'}	11.0	10.7	3.3	3.3	11.7	11.1	3.0		2.7	3.0	11.6	10.5
J _{3',4'}	9.1		2.2	1.9	3.2	2.5	3.0		2.8	3.0	3.2	2.8
J4',5'	9.9		5.1	2.8	<1		6.8		1.4	10.6 ^a	2.5 ^b	2.0 ^c

Table 5. Selected ¹H NMR coupling constants (Hz) of the carbohydrate frame for 2'-Deoxy-2'-phenylselenenyl-pyranosyl nucleosides 4a-b, 5a-b, 7a-b, 8a-b, 9a-b and 10a-b.

^a J_{4',5"=6.1} Hz. ^b J_{4',5"=1.1} Hz. ^c J_{4',5"=0} Hz

benzyl-D-galactal (2b) was excellent in ether as well as in acetonitrile, in both cases compound 7b being the main one obtained (Table 4), which originated from the selenonium ion formed by below the plane attack of the selenium reagent.

The reaction of 3,4-di-O-acetyl-D-arabinal (3a) with PhSeCl, AgOTf and bis-(trimethylsilyl)uracil gave very low selectivity in ether as well as in acetonitrile, producing a 1:1 mixture of compounds 9a and 10a. A small amount of product originated by S_N2' reaction was also detected in this case. Similar results were obtained when starting from 3,4-di-O-benzyl-D-arabinal (3b) and using acetonitrile as the solvent; however, in ether product 10b was obtained with a selectivity of 92%. In this case the main product arose from above the plane attack of the electrophilic part of PhSeCl to arabinal, that is, opposite to the ring substituents.

Structure assignment

The structure of the 2'-deoxy-2'-phenylselenenyl nucleosides was established by ¹H and ¹³C NMR spectroscopy on the basis of the following facts: (1) presence of the uracil in the products, which wasconfirmed by the double bond signals in the ¹H NMR spectra (6.70-7.00 ppm, d, H-6; 5.00-5.50 ppm, dd, H-5) and in the ¹³C NMR spectra (~140 ppm, C-6; ~102 ppm, C-5); (2) introduction of the phenylselenenyl group was proved by the presence in the ¹³C NMR spectra of a signal at 43-50 ppm, assigned to C-2', typical of carbon bonded to selenium; (3) coupling constants $J_{1,2}^{1}$, $J_{2,3}^{2}$, $J_{3,4}^{2}$ and $J_{4,5}^{2}$ are indicative of configuration of positions 1 and 2, and also of the preferred conformation of the sugar ring. Thus, as can be seen in the Table 5, a J_{1:2}>9 Hz for all the obtained compounds, except for 9b, indicates a trans-diaxial arrangement for protons H-1'and H-2' which implies a trans-equatorial disposition for uracil and selenium. The value of $J_{2',3'}$ is also higher than 9 Hz for compounds 4a-b, 7a-b and 10a-b, confirming a trans-diaxial arrangement for protons H-1', H-2' and H-3'. These facts, together with the values of the other coupling constants, corroborates a ßgluco and β -galacto configuration with a ${}^{4}C_{1}$ conformation 15 for compounds 4a-b and 7a-b, respectively. In the case of arabino derivatives 10a-b the small size of both J4',5' and J4',5' indicates a ¹C4 conformation for these compounds. On the other hand, a J_{2',3} = 3-4 Hz for compounds 5a-b and 8a-b corresponds to an axialequatorial coupling; as H-2' is always axial, H-3' must be equatorial, which supposes that a chair inversion has taken place and that the conformation ${}^{1}C_{4}$ preponderates in these compounds. This fact is confirmed by the low value of $J_{3',4'}$ and $J_{4',5'}$. Compound 9a shows $J_{2',3'}$ and $J_{3',4'} \approx 2.8$ Hz which, since $H_{2'}$ is axial, indicates that $H_{3'}$ is equatorial and consequently $H_{4'}$ is axial, which confirms a 4C_1 conformation for this compound.

12225

Compound **9b** is the only case where $J_{1',2'}$ is small, suggesting that uracil and the PhSe group are not in a *trans* diequatorial disposition, as in all the other compounds. A value of $J_{4',5'}=10.1$ Hz indicates that $H_{4'}$ and $H_{5'}$ are axial, but the other coupling constants are very small, which implies that the other substituents are in axial or pseudo-axial disposition. It must correspond with a twist-boat conformation probably trying to avoid the steric interactions among the bulky substituents at C-2, C-3 and C-4.

The next NMR spectroscopic data confirms the structure of compound 6: (1) the absence of aromatic protons and the signal between 40 and 50 ppm in the ¹³C NMR spectrum, showing that no PhSe group exists, (2) the presence of only two acetate group signals and (3) the signals at 6.55 ppm and 6.18 ppm in the ¹H NMR spectrum and at 132.8 ppm and 127.2 ppm in the ¹³C NMR spectrum, characteristic of a double bond. These values are in good agreement with those for other similar compounds described in the literature.^{7e,8}

DISCUSSION

It is well known that NIS,¹¹⁻¹³ PhSCl⁹ and PhSeCl¹⁰ mediated O-glycosylation reaction gives exclusively alkyl 2-deoxy-2-X-nucleosides (X = I, SPh or SePh) having X and alkoxy group *trans* disposed. The results are explained in terms of a two step mechanism (Scheme 1), consisting of electrophilic addition of NIS, PhSCl or PhSeCl to the glycal double bond to give a cyclic cation, followed by a regioselective nucleophilic attack by an alcohol at C-1.

In our case the selenium-mediated synthesis of nucleosides from glycals does not take place at room temperature because the bis-(trimethylsilyl)uracil is not nucleophilic enough, therefore the presence of a chlorine activator is necessary to obtain the corresponding nucleosides.

It has been recently shown¹² that the stereochemistry of NIS-alcohol addition to glycals is determined at the electrophilic step (iodine addition) or at the nuclephilic step (alcohol attack) depending on the solvent. In general, in ether type solvents an isomerization between iodonium cations takes place, while in acetonitrile the iodonium cation formation is irreversible, the stereoisomer distribution reflecting in this last case the ratio of iodonium cations.

On the other hand, in the selenium mediated synthesis of O-glycosides from glycals an isomerization process between selenonium cations has been proposed to explain the distribution products obtained.^{10a}

The results collected in Table 4 show that the stereochemistry is dependent on the solvent and on the glycal structure, but also on the protecting groups.

The solvent has an influence not only on the selectivity but also on the reaction rate, being faster in ether than in acetonitrile. Concerning the stereoselectivity, compounds arising from the less hindered selenonium intermediate, [A] for glucal and galactal derivatives and [B] for arabinal derivatives (Scheme 1), were favoured when ether was the solvent, while in acetonitrile the percentage of products derived from the intermediate [B] increased. This fact agrees with the Horton¹² proposal, that an isomerization process between intermediates [A] and [B] takes place, being faster in ether than in acetonitrile, and also with the observations of Beau^{10a} that phenylselenonium cations isomerize easily, expecially in the presence of Lewis acids. In this way, it must be taken into consideration that TMSOTf is probably generated in the reaction medium in the PhSeCI-mediated synthesis of nucleosides from glycals. The different stereoselectivities observed for different selenium reagents (Table 3) confirms the existence of an isomerization process between the selenonium cation and the 1-halo-2-phenylselenenylpyranose and also between selenonium cations [A] and [B] (Scheme 1).

Protecting groups and glycal structure play a determining role in the control of stereoselectivity. The preferred conformation for D-glucal and especially for D-galactal and D-arabinal derivatives¹⁶ is ⁴H₅ (Table 4), and although differences in stereoselectivity are observed when starting from different glycals, the preferred

side of attack to the double bond of the glycal in the more stable conformation, or the existence of an isomerization process can not exclusively explain these differences. In Table 4 it can be seen that stereoselectivity is strongly dependent on the type of protecting group. Thus, for the acetylated glycal 2a a relatively higher percentage of α -talo derivative is obtained, indicating that the more hindered selenonium cation is principally formed. It appears that electrostatic effects might be responsible for the stereochemistry of the first step.¹⁷ This effect is less important in the glycals 1a and 3a, given that the dipole moment of ring oxygen and the C-4 acetoxy group are opposite, and so the net dipole moment should be smaller, which would explain the lower stereoselectivity obtained in these cases.

On the contrary, the results from benzylated glycals 1b, 2b and 3b clearly show that steric factors exert the major influence in these cases, given that phenylselenenyl nucleosides originated by the opening of the less hindered selenonium cation ([A] for glucal and galactal derivatives and [B] for arabinal derivatives) are principally obtained. In these cases the stereoselectivity is better in ether than in acetonitrile, as this corresponds to a faster equilibrium process in which the more stable cyclic cation intermediate is the major one.

In summary, the face selectivity in the reaction of glycals with bis-(trimethylsilyl)uracil induced by PhSeCl is determined by the protecting groups present in the glycal, the stereochemistry of glycal substituents, the solvent and also by the activating reagent. The reaction takes place in two steps, and the stereoselectivity depends on the preferred formation of the cyclic phenylselenonium cation intermediate by the more or less hindered face. Glycal structure and protecting groups determine which selenonium cation is preferentially formed. The more hindered intermediate [B] (Scheme 1) principally results from acetylated glycals and especially when the acetoxy group at position 4 is axial, and the less hindered intermediate [A] is preferentially formed starting from benzyl protected glycals and using PhSeCl and ether. The solvent, in addition to the different stabilisations of conformers, influences the stereoselectivity by increasing the isomerization of selenonium cations, leading to a thermodynamically more stable selenonium cation.

2'-DEOXYPYRANOSYL NUCLEOSIDES

As can be seen in Table 4, benzyl glycals gave better stereoselectivities in β -gluco, β -galacto and α arabino 2'-deoxy-2'-phenylselenenyl-pyranosyl nucleosides. Compounds 7b and 10b, which were obtained with the higher stereoselectivity, and their corresponding acetyl derivatives 7a and 10a were treated with tributyltin hydride and AIBN in benzene and heated to reflux to give the 2'-deoxy-pyranosyl nucleosides 13b-14b and 13a-14a respectively, in yields above 80%.



As has been described before, tri-O-benzyl derivatives 4b and 5b were obtained as an inseparable mixture. When, a 4:1 mixture of these compounds was treated with tributyltin hydride 2-deoxy-pyranosyl

Comp./J	H6	H5	H1'	H2'a	H2'e	Н3'	H4'	H5'	H6'	H6"
11a	7.30	5.76	5.85	1.80	2.40	5.14	4.98	3.80	4.23	4.04
11b	7.4-7.1	5.76	5.69	1.65	2.45	3.84		3.42		
12	7.5-7.2	5.70	6.07	2.19	-2.01	3.8	3.53	4.30	3.9-3.8	3.70
13a	7.43	5.83	5.86	2.15	5-2.10	5.17	5.36		4.814.01	
13b	7.43	5.66	5.64	2.10)-2.00	3.64	3.82		3.64	
14 a	7.42	5.80	6.14	2.30)-2.00	5.47	5.24	4.39	4.65	4.29
14b	7.51	5.78	5.61	2.24	-2.03	3.71	3.71		4.20	3.48

Table 6. Selected ¹H NMR chemical shifts for compounds 11a-b, 12, 13a-b and 14a-b.

Table 7. ¹H NMR coupling constants for compounds 11a-b, 12, 13a-b and 14a-b.

Comp./J	J _{5,6}	J _{1',2'a}	J _{1',2'e}	J _{2'a,2'e}	J _{2'a,3'}	J _{2'e,3}	J _{3',4'}	J _{4',5'}	J _{5',6'}	J _{5',6"}	J6',6"
11a	8.1	11.1	2.0	12.3	11.1	5.1	10.9	9.7	5.1	2.0	12.5
11b	8.3	11.1	2.2	11.1	11.1	4.9					
12	8.0	10.1	3.6				3.0	3.0		5.5	10.1
13a	8.2	13.0	3.3		11.5	5.6	3.0			**	
13b	8.1	8.0	5.0								
14 a	8.2	9.1	3.7		3.3	3.3	3.3	5.6	8.8	2.9	12.1
14b	8.2	10.9	2.7					1.7	12.8ª		

a J5',5".

nucleosides 11b and 12 were also obtained in the same ratio as an inseparable mixture. A small fraction of this mixture was purified by HPLC for identification purposes. In the same way 11a was obtained from 4a.

The replacement of the PhSe group by a proton does not alter the conformation, conserving, in the obtained 2'-deoxynucleosides, the same ${}^{4}C_{1}$ or ${}^{1}C_{4}$ conformation that they had in the starting 2'-deoxy-2'-phenylselenenyl nucleoside.^{3,7a,18,19} The value of the coupling constants, $J_{1',2a'} > 9Hz$ and $J_{1',2c'} > 4Hz$ (Table 7), confirms the equatorial disposition for the uracil in all the 2'-deoxypyranosyl nucleosides synthesised.

In conclusion, 2'-deoxy-2'-phenylselenenyl pyranosyl nucleosides have been stereoselectively obtained from glycals and converted into the corresponding 2'-deoxypyranosyl nucleosides in good yields.

EXPERIMENTAL SECTION

General Procedures: Melting points were measured in a Büchi 510 apparatus and appear uncorrected. Optical rotations were measured at room temperature in 10 cm cells in a Perkin-Elmer 241 polarimeter. ¹H NMR and ¹³C NMR spectra were recorded in a Varian Gemini 300 MHz (300 and 75.4 MHz resp.) apparatus, with CDCl₃ as solvent and using Me₄Si (δ =0)and the central solvent peak at ∂ 77.0 ppm respectively as internal reference. Elemental analyses were determined using a Carlo-Erba Microanalysis. Flash column chromatography was performed with silica gel 60 A CC (230-400 mesh). TLC plates were prepared by using Kieselgel 60 PF₂₅₄ (E. Merk). HPLC was performed with a C-18 silicagel column (25mm x 10 cm) using acetonitrile/water 60:40 as eluent. Solvents for chromatography were distilled at atmospheric pressure prior to use. Dichloromethane was distilled from P₂O₅. Benzene was dried by distillation, under nitrogen, from sodium benzophenone ketyl. Other solvents were purified and dried by using standard procedures. All the reactions were carried out under an argon atmosphere using standard syringe techniques.

General Procedure for the synthesis of 2'-phenylselenenyl-pyranosyl nucleosides from glycals. To a solution of glycal (0.25 mmol) in 1 ml of dry solvent, keeped under argon and light protected, 0.37 mmol of the phenylselenenyl reagent was added at room temperature, after 5 minutes 0.5 mmol of bis-(trimethylsilyl) uracil and finally 0.42 mmol of the catalyst were also added to the reaction flask. After the reaction was finished (about 0.5-24 h) ethyl acetate (25 mL) were added and the reaction mixture was filtered off through a silicagel-Celite pad (the Celite pad was washed with 2x10 mL of ethyl acetate). Evaporation of the solvent gave the crude product mixture which was subjected to TLC or flash chromatography.

 $1-(2'-Deoxy-2'-phenylselenenyl-3',4',6'-tri-O-acetyl-B-D-gluco-pyranosyl)-uracil (4a) and <math>1-(2'-deoxy-2'-phenylselenenyl-3',4',6'-tri-O-acetyl-\alpha-D-manno-pyranosyl)-uracil (5a).$

The general procedure was applied starting from 3,4,6-tri-O-acetyl-D-glucal, phenylselenenyl chloride, silver triflate and bis-(trimethylsilyl)uracil in ether. The reaction was stopped after 0.5 hours. Flash chromatography (ethyl acetate-hexane 3:2) of the reaction crude gave 59 mg (44%) of 4a and 44 mg of a mixture of 5a/6 (2:1). A small sample of this mixture was purified by HPLC for characterisation purposes.

(4a). Mp= 98-99°C, $[\alpha]_D^{20}$ +69.2° (c 0.5, CHCl₃). ¹H NMR: δ 9.45 (s, 1H, NH); 7.50-7.10 (Ph); 6.88 (d,1H, J_{H-6,H-5}=8.2 Hz, H-6); 5.94 (d, 1H, J_{H-1',H-2}=11.0 Hz, H-1'); 5.39 (dd, 1H, J_{H-5,NH}=1.8 Hz, H-5); 5.25 (dd, 1H, J_{H-3',H-4}=9.1 Hz, J_{H-3',H-2}=11.0 Hz, H-3'); 5.05 (dd, 1H, J_{H-4',H-5'}=9.9 Hz, H-4'); 4.22 (dd, 1H, J_{H-6',H-5'}=12.6 Hz, J_{H-6',H-5}=4.8 Hz, H-6'); 4.02 (dd, 1H, J_{H-5',H-5'}=2.0 Hz, H-6''); 3.85 (ddd, 1H, H-5'); 3.28 (t, 1H, H-2'); 2.07 (Me); 2.01 (Me); 2.01 (Me). ¹³C NMR: δ 170.8 (CH₃<u>C</u>O); 170.1 (CH₃<u>C</u>O); 170.0 (CH₃<u>C</u>O); 162.8 (C-4); 150.4 (C-2); 138.0-129.1 (Ph); 136.6 (C-6); 103.1 (C-5); 82.5 (C-1'); 74.4 (C-4'); 72.4 (C-3'); 68.8 (C-5'); 61.7 (C-6'); 48.3 (C-2'); 20.5 (2xMe); 20.4 (Me). IR: 1748, 1737, 1735 cm⁻¹ (\sqrt{CO}); 1691, 1687 cm⁻¹ (\sqrt{CO}); 1632 cm⁻¹ ($\sqrt{CH=CH}$). Anal. Calcd for C₂₂H₂₄N₂O9Se: C, 49.01; H, 4.45; N, 5.19. Found: C, 48.62; H, 4.52; N, 5.10.

(5a). Mp = 74 - 76°C, $[\alpha]_D^{20}$ -41.7° (c 0.2, CHCl₃). ¹H NMR: δ 8.47 (s, 1H, NH); 7.45-7.10 (Ph); 6.93 (d,1H, J_{H-6,H-5}=8.2 Hz, H-6); 6.06 (d, 1H, J_{H-1',H-2'}=10.8 Hz, H-1'); 5.49 (dd, 1H, J_{H-3',H-4}=2.2 Hz, J_{H-3',H-2'}=3.3 Hz, H-3'); 5.35 (dd, 1H, J_{H-5,NH}=2.2 Hz, H-5); 4.81 (dd, 1H, J_{H-4',H-5'}=5.1 Hz, H-4'); 4.52 (dd, 1H, J_{H-6',H-5'}=11.8 Hz, J_{H-6',H-5'}=7.9 Hz, H-6'); 4.29 (m, 1H, H-5'); 4.18 (dd, 1H, J_{H-6',H-5'}=5.0 Hz, H-6''); 3.87

(dd, 1H, H-2'); 2.14 (Me); 2.10 (Me); 2.00 (Me). ¹³C NMR: δ 170.6 (CH₃<u>C</u>O); 169.0 (CH₃<u>C</u>O); 168.9 (CH₃<u>C</u>O); 162.4 (C-4); 150.0 (C-2); 140.2 (C-6); 138.0-128.9 (Ph); 102.4 (C-5); 80.1 (C-1'); 75.3 (C-4'); 71.6 (C-3'); 67.0 (C-5'); 60.2 (C-6'); 43.2 (C-2'); 20.9 (Me); 20.6 (2xMe). IR: 1744, 1717 cm⁻¹ (\sqrt{CO}); 1695, 1685 cm⁻¹ (\sqrt{CO} , anillo); 1620 cm⁻¹ ($\sqrt{CH=CH}$). Anal. Calcd for C₂₂H₂₄N₂O₉Se: C, 49.01; H, 4.45; N, 5.19. Found: C, 48.75; H, 4.56; N, 5.05.

(6). Mp = 184 - 185°C, $[\alpha]_D^{21}$ +46.5° (*c* 1, CHCl₃). ¹H NMR: δ 8.85 (s, 1H, NH); 7.20 (d, 1H, J_{H-6,H-5}= 8.1 Hz, H-6); 6.55 (dd, 1H, J_{H-1',H-2}= 2.0 Hz, J_{H-1',H-3}= 3.9 Hz, H-1'); 6.18 (dt, 1H, J_{H-2',H-3}= 10.3 Hz, J_{H-2',H-4}= 2.0 Hz, H-2'); 5.80-5.75 (m, 2H, H-5, H-3'); 5.38 (dt, 1H, J_{H-4',H-3}= J_{H-4',H-2}= 2.0 Hz, J_{H-4',H-5}= 9.1 Hz, H-4'); 4.25-4.17 (m, 2H, H-6', H-6''); 4.03 (m, 1H, H-5'); 2.12 (Me); 2.09 (Me). ¹³C NMR: δ 170.9 (CH₃<u>C</u>O); 170.2 (CH₃<u>C</u>O); 162.5 (C-4); 150.1 (C-2); 140.0 (C-6); 132.8 (C-2'); 127.2 (C-3'); 103.5 (C-5); 78.6 (C-1'); 75.1 (C-4'); 64.0 (C-5'); 62.6 (C-6'); 20.9 (Me); 20.8 (Me). Anal. Calcd for C₁₄H₁₆N₂O₇: C, 51.88; H, 4.93; N, 8.64. Found: C, 51.52; H, 4.83; N, 8.40.

1-(2'-Deoxy-2'-phenylselenenyl-3',4',6'-tri-O-benzyl- β -D-gluco-pyranosyl)-uracil (4b) and 1-(2'-deoxy-2'-phenylselenenyl-3',4',6'-tri-O-benzyl- α -D-manno-pyranosyl)-uracil (5b).

Glycosylation was carried out in ether starting from 3,4,6-tri-O-benzyl-D-glucal and phenylselenenyl chloride, bis-(trimethylsilyl)uracil and silver triflate in ether. The reaction was stopped after 0.5 hours. Flash chromatography (ethyl acetate-hexane 2:3) did not allow the separation of the phenylselenenyl nucleosides, obtaining 144 mg (85%) of a 4:1 mixture of compound **4b** and **5b**. A small sample of this mixture was purified by HPLC for identification purposes.

(4b). ¹H NMR: δ 8.40 (s, 1H, NH); 7.50-7.10 (Ph); 6.79 (d, 1H, J_{H-6,H-5}=7.9 Hz, H-6); 5.88 (d, 1H, J_{H-1',H-2'}=10.7 Hz, H-1'); 5.23 (dd, 1H, J_{H-5,NH}=2.2 Hz, H-5); 4.91 (d, 1H, J_{gem}=10.1 Hz, CH₂Ph); 4.83 (d, 1H, J_{gem}=10.1 Hz, CH₂Ph); 4.78 (d, 1H, J_{gem}=12.8 Hz, CH₂Ph); 4.55 (d, 1H, J_{gem}=12.8 Hz, CH₂Ph); 4.46 (d, 1H, J_{gem}=12.1 Hz, CH₂Ph); 4.39 (d, 1H, J_{gem}=12.1 Hz, CH₂Ph); 3.74-3.50 (m, 5H, H-3', H-4', H-5', H-6', H-6''); 3.23 (t, 1H, J_{H-2',H-3'}=10.7 Hz, H-2'). ¹³C NMR: δ 162.0 (C-4); 150.0 (C-2); 138.9 (C-6); 139.3-128.1 (Ph); 102.5 (C-5); 82.6 (C-1'); 82.9 (C-4'); 79.0 (C-3'); 77.8 (CH₂Ph); 77.8 (CH₂Ph); 75.6 (CH₂Ph); 73.5 (C-5'); 68.3 (C-6'); 51.4 (C-2'). IR: 1716, 1696 cm⁻¹ (\sqrt{CO}); 1635 cm⁻¹ ($\sqrt{CH=CH}$).

(5b). ¹H NMR: δ 7.80 (s, 1H, NH); 7.40-7.10 (Ph); 6.77 (d, 1H, J_{H-6,H-5}=8.2 Hz, H-6); 6.19 (d, 1H, J_{H-1',H-2}=10.9 Hz, H-1'); 5.04 (dd, 1H, J_{H-5,NH}=2.1 Hz, H-5); 4.60-4.25 (m, 7H, 3xC<u>H2</u>Ph, H-5'); 4.16 (dd, 1H, J_{H-3',H-2}=3.3 Hz, J_{H-3',H-4}=1.9 Hz, H-3'); 3.78 (dd, 1H, J_{H-6',H-6}=10.1 Hz, J_{H-6',H-5}=7.1 Hz, H-6'); 3.69 (dd, 1H, H-2'); 3.59 (dd, 1H, J_{H-4',H-5}=2.8 Hz, H-4'); 3.64 (dd, 1H, J_{H-6',H-5}=6.8, H-6''). RMN ¹³C NMR: δ 162.0 (C-4); 150.0 (C-2); 139.5 (C-6); 138.0-127.7 (Ph); 101.8 (C-5); 78.7 (C-4'); 77.3 (C-1'); 76.2 (C-3'); 73.4 (<u>C</u>H₂Ph); 73.1 (<u>C</u>H₂Ph); 71.1 (<u>C</u>H₂Ph); 71.1 (C-5'); 67.3 (C-6'); 46.6 (C-2'). IR: 1712, 1698 cm⁻¹ (\sqrt{CO}); 1632 cm⁻¹ ($\sqrt{CH=CH}$).

1-(2'-Deoxy-2'-phenylselenenyl-3',4',6'-tri-O-acetyl-β-D-galacto-pyranosyl)-uracil (7a) and 1-(2'-deoxy-2'-phenylselenenyl-3',4',6'-tri-O-acetyl-α-D-talo-pyranosyl)-uracil (8a).

Following the general procedure the glycosylation was performed starting from 3,4,6-tri-O-acetyl-D-galactal and using phenylselenenyl chloride, bis-(trimethylsilyl)uracil and silver triflate in ether as the solvent for 4 hours. Thin layer chromatography (ethyl acetate-hexane (1:3) of the reaction crude gave 54 mg of 7a and 54 mg of 8a, total yield 80% (7a/8a = 50:50).

(7a). Mp = 104-106°C, $[\alpha]_D^{20}$ +37.5° (c 0.3, CHCl3). ¹H NMR: δ 8.95 (s, 1H, NH); 7.60-7.20 (Ph); 7.10 (d, 1H, J_{H-6,H-5}=8.3 Hz, H-6); 5.96 (d, 1H, J_{H-1',H-2}=10.7 Hz, H-1'); 5.61 (dd, 1H, J_{H-5,NH}=2.0 Hz, H-5);

5.37 (d, 1H, $J_{H-3',H-4}=3.2$ Hz, H-4'); 4.91 (dd, 1H, $J_{H-3',H-2}=11.7$ Hz, H-3'); 4.24-3.96 (m, 3H, H-5', H-6', H-6''); 3.40 (dd, 1H, H-2'); 2.19 (Me); 2.08 (Me); 2.03 (Me). ¹³C NMR: δ 162.6 (3xCH₃CO); 162.4 (C-4); 150.2 (C-2); 138.6 (C-6); 138.0-129.7 (Ph); 103.3 (C-5); 82.5 (C-1'); 73.3 (C-4'); 70.3 (C-3'); 66.5 (C-5'); 61.4 (C-6'); 43.6 (C-2'); 20.5 (3xMe). IR: 1748, 1735 cm⁻¹ (\sqrt{CO}); 1697, 1690 cm⁻¹ (\sqrt{CO}); 1630 cm⁻¹ ($\sqrt{CH=CH}$). Anal. Calcd for C₂₂H₂₄N₂O9Se: C, 49.01; H, 4.45; N: 5.19. Found: C, 48.90; H, 4.57; N, 5.02. (8a). Mp= 114-116 °C, $[\alpha]_D^{20}$ -19.0° (c 0.6, CHCl₃). ¹H NMR: δ 8.53 (s, 1H, NH); 7.50-7.20 (Ph); 7.03 (d, 1H, J_{H-6,H-5}=8.2 Hz, H-6); 6.16 (d, 1H, J_{H-1',H-2'}=10.7 Hz, H-1'); 5.93 (t, 1H, J_{H-3',H-2}=J_{H-3',H-4}=3.0 Hz, H-3'); 5.44 (dd, 1H, J_{H-5,NH}=1.7 Hz, H-5); 5.31 (dd, 1H, J_{H-4',H-5'}=6.8 Hz, H-4'); 4.91 (dd, 1H, J_{H-6',H-6'}=12.8 Hz, J_{H-6',H-5'}=10.0 Hz, H-6'); 4.45-4.36 (m, 1H, H-5'); 4.25 (dd, 1H, J_{H-6',H-5'}=2.2 Hz, H-6''); 3.84 (dd, 1H, H-2'); 2.23 (Me); 2.08 (Me); 2.06 (Me). ¹³C NMR: δ 172.0 (CH₃CO); 162.6 (C-4); 160.6 (2xCH₃CO); 150.0 (C-2); 140.5 (C-6); 135.5-127.0 (Ph); 102.6 (C-5); 79.3 (C-1'); 74.0 (C-4'); 70.2 (C-3'); 67.1 (C-5'); 59.3 (C-6'); 45.3 (C-2'); 20.6 (Me); 20.6 (Me); 20.4 (Me). Anal. Calcd for C₂₂H₂₄N₂O9Se: C, 49.01; H, 4.45; N, 5.19. Found: C, 48.61; H, 4.54; N, 4.99.

1-(2'-Deoxy-2'-phenylselenenyl-3',4',6'-tri-O-benzyl-β-D-galacto-pyranosyl)-uracil (7b) and 1-(2'-deoxy-2'-phenylselenenyl-3',4',6'-tri-O-benzyl-α-D-talo-pyranosyl)-uracil (8b).

The general procedure was applied to 3,4,6-tri-O-benzyl-D-galactal and phenylselenenyl chloride, bis-(trimethylsilyl)uracil and silver triflate, in ether as the solvent. The reaction was stopped after 1 hour. Thin layer chromatography (ethyl acetate-hexane 1:1) of the reaction crude gave 128 mg of 7b and 13 mg of 8b, total yield 82% (7b/8b = 91:9).

(7b). M.p. 132-134 °C; $[\alpha]_{D^{20}}$ +74.3° (*c* 0.45, CHCl₃). ¹H NMR: δ 8.27 (s, 1H, NH); 7.60-7.15 (m, 20 H, Ph); 7.00 (d, 1H, J_{H-6,H-5}= 8.2 Hz, H-6); 5.87 (d, 1H, J_{H-1',H-2'}= 10.6 Hz, H-1'); 5.39 (dd, 1H, J_{H-5,NH}= 1.9 Hz, H-5); 4.90 (d, 1H, J_{gem}= 11.3 Hz, CH₂Ph); 4.75 (d, 1H, J_{gem}= 11.3 Hz, CH₂Ph); 4.59 (d, 1H, J_{gem}= 11.3 Hz, CH₂Ph); 4.57 (d, 1H, J_{gem}= 11.3 Hz, CH₂Ph); 4.46 (d, 1H, J_{gem}= 11.8 Hz, CH₂Ph); 4.41 (d, 1H, J_{gem}= 11.8 Hz, CH₂Ph); 4.50 (m, 1H, H-4'); 3.70 (dd, 1H, J_{H-2',H-3'}= 11.1 Hz, H-2'); 3.68 (m, 1H, H-5'); 3.56 (dd, 1H, J_{H-5',H-6''}= 9.0 Hz, J_{H-6',H-5'}= 7.3 Hz, H-6'); 3.50 (dd, 1H, J_{H-6',H-5'}= 5.7 Hz, H-6''); 3.44 (dd, 1 H, J_{H-3',H-4'}= 2.5 Hz, H-3'). ¹³C NMR: δ 162.39 (C-4); 150.00 (C-2); 139.48 (C-6); 135.24-127.36 (Ph); 102.62 (C-5); 83.19 (C-1'); 79.89 (C-4'); 75.66 (C-3'); 74.84 (CH₂Ph); 73.52 (CH₂Ph); 72.37 (CH₂Ph); 72.07 (C-5'), 67.94 (C-6'); 47.27 (C-2'). Anal. Calcd. for C₃₇H₃₆N₂O₆Se: C, 65.03; H, 5.26; N, 4.09. Found: C, 64.64; H, 5.35; N, 4.22.

(8b). ¹H NMR: δ 8.03 (s, 1H, NH); 7.50-7.20 (m, 21 H, Ph, H-6); 5.73 (dd, 1H, $J_{H-5,H-6}$ = 8.0 Hz, $J_{H-5,NH}$ = 1.7 Hz, H-5); 5.68 (d, 1H, $J_{H-1',H-2'}$ = 9.0 Hz, H-1'); 4.94 (d, 1H, J_{gem} = 11.4 Hz, CH2Ph); 4.60 (d, 1H, J_{gem} = 11.4 Hz, CH2Ph); 4.60 (d, 1H, J_{gem} = 11.4 Hz, CH2Ph); 4.60 (s, 2H, CH2Ph); 4.50 (d, 1H, J_{gem} = 11.9 Hz, CH2Ph); 4.44 (d, 1H, J_{gem} = 11.9 Hz, CH2Ph); 3.89 (m, 1H, H-3'); 3.74-3.60 (m, 3H, H-4', H-5', H-2'); 3.59-3.55 (m, 2H, H-6', H-6''). ¹³C NMR: δ 140.10 (C-6); 128.56-127.34 (Ph); 102.76 (C-5); 19.90 (C-1'); 77.21 (C-4'); 76.20 (C-3'); 74.54 (CH2Ph); 73.56 (CH2Ph); 71.61 (CH2Ph); 70.58 (C-5'); 68.65 (C-6'); 41.00 (C-2').

1-(2'-Deoxy-2'-phenylselenenyl-3',4'-di-O-acetyl-8-D-*ribo*-pyranosyl)-uracil (9a) and 1-(2'-deoxy-2'-phenylselenenyl-3',4'-di-O-acetyl-α-D-*arabino*-pyranosyl)-uracil (10a).

Glycosylation was carried out starting from 3,4-di-O-acetyl-D-arabinal in ether and using phenylselenenyl chloride, bis-(trimethylsilyl)uracil and silver triflate. After 2 hours the reaction was stopped. Purification of the reaction crude by thin layer chromatography, successive elution with methylene chloride/hexane/methanol 10:6:0.2, gave 35 mg of **9a** and 52 mg of **10a** as an oil, total yield **75%** (**9a/10a** = 40:60).

(9a). $[\alpha]_D^{20}$ +36.7° (c 0.4, CHCl₃). ¹H NMR: δ 8.22 (s, 1H, NH); 7.40-7.10 (Ph); 6.76 (d, 1H, J_{H-6,H-5}=8.2 Hz, H-6); 6.03 (d, 1H, J_{H-1',H-2}=10.7 Hz, H-1'); 5.89 (dd, 1H, J_{H-3',H-2}=2.7 Hz, J_{H-3',H-4}=2.8 Hz, H-3'); 5.26 (dd, 1H, J_{H-5,NH}=2.4 Hz, H-5); 5.01-4.95 (m, 2H, H-4', H-5'); 3.86 (dd, 1H, J_{H-5',H-5}=9.1 Hz, J_{H-5',H-4}=1.4 Hz, H-5''); 3.37 (dd, 1H, H-2'); 2.15 (Me); 1.95 (Me). ¹³C NMR: δ 169.7 (2xCH₃<u>C</u>O); 161.7 (C-4); 150.0 (C-2); 139.0 (C-6); 135.4-126.6 (Ph); 102.7 (C-5); 81.9 (C-1'); 70.4 (C-4'); 66.8 (C-3'); 63.8 (C-5'); 46.7 (C-2'); 20.6 (2xMe). Anal. Calcd. for C₁₉H₂₀N₂O₇Se: C, 48.82; H, 4.28; N, 5.99. Found: C, 48.25; H, 4.32; N, 5.81.

(10a). $[\alpha]_D^{20}$ -25.0° (c 0.52, CHCl₃) .¹H NMR: δ 8.40 (s, 1H, NH); 7.50-7.10 (Ph); 7.01 (d, 1H, J_{H-6,H-5}=8.3 Hz, H-6); 5.74 (d, 1H, J_{H-1',H-2}=10.7 Hz, H-1'); 5.49 (dd, 1H, J_{H-5,NH}=2.4 Hz, H-5); 5.21-5.18 (m, 1H, H-4'); 4.86 (dd, 1H, J_{H-3',H-2}=11.6 Hz, J_{H-3',H-4}=3.2 Hz, H-3'); 3.98 (dd, 1H, J_{H-5',H-5}=13.4 Hz, J_{H-5',H-4}=2.5 Hz, H-5'); 3.69 (dd, 1H, J_{H-5',H-4}=1.1 Hz, H-5''); 3.43 (dd, 1H, H-2'); 2.10 (Me); 2.03 (Me). ¹³C NMR: δ 169.9 (CH₃<u>C</u>O); 169.0 (CH₃<u>C</u>O); 162.0 (C-4); 150.0 (C-2); 139.0 (C-6); 136.2-129.2 (Ph); 103.0 (C-5); 83.4 (C-1'); 70.1 (C-4'); 67.5 (C-3'); 67.0 (C-5'); 44.3 (C-2'); 21.0 (Me); 20.7 (Me). Anal. Calcd. for C₁₉H₂₀N₂O₇Se: C, 48.82; H, 4.28; N, 5.99. Found: C, 48.37; H, 4.36; N, 5.85.

1-(2'-Deoxy-2'-phenylselenenyl-3',4'-di-O-benzyl-β-D-*ribo*-pyranosyl)-uracil (9b) and 1-(2'-deoxy-2'-phenylselenenyl-3',4'-di-O-benzyl-α-D-*arabino*-pyranosyl)-uracil (10b).

Following the general procedure the glycosylation was carried out starting from 3,4-di-O-benzyl-D-arabinal in ether and using phenylselenenyl chloride, bis-(trimethylsilyl)uracil and silver triflate. After 1 hours the standard work-up and purification by thin layer chromatography (ethyl acetate-hexane 1:1) of the reaction crude gave 92 mg of 10b and 8 mg of 9b; total yield 71% (9b/10b = 8:92).

(10b). M.p. 69-70 °C, $[\alpha]_D^{21}$ -74.0° (*c* 0.5, CHCl₃), ¹H NMR: δ 9.04 (s, 1H, NH); 7.40-7.15 (m, 10 H, Ph); 7.08 (d, 1H, J_{H-6,H-5} = 8.1 Hz, H-6); 5.82 (d, 1H, J_{H-1',H-2'} = 10.5 Hz, H-1'); 5.43 (dd, 1H, J_{H-5,NH} = 2.1 Hz, H-5); 4.74 (d, 1H, J_{gem}= 12.2 Hz, CH2Ph); 4.65 (d, 1H, J_{gem}= 12.2 Hz, CH2Ph); 4.62 (d, 1H, J_{gem}= 11.5 Hz, CH2Ph); 4.51 (d, 1H, J_{gem}= 11.5 Hz, CH2Ph); 4.12 (dd, 1H, J_{H-5',H-5'} = 13.0 Hz, J_{H-5',H-4} = 2.0 Hz, H-5'), 3.75 (t, 1H, J_{H-2',H-3'} = 10.5 Hz, H-2'); 3.74 (m, 1H, H-4'); 3.43 (d, 1H, H-5''), 3.41 (dd, 1H, J_{H-3',H-4} = 2.8 Hz, H-3'). ¹³C NMR: δ 163.09 (C-4); 150.81 (C-2); 139.77 (C-6); 135.25-127.08 (Ph); 103.16 (C-5); 83.97 (C-1'); 78.64 (C-4'); 72.11 (CH2Ph); 71.86 (CH2Ph); 71.38 (C-3'); 66.36 (C-5'), 47.29 (C-2'). Anal. Calcd for C₂₉H₂₈N₂O₅Se: C, 61.84; H, 4.97; N, 4.97. Found: C, 61.45; H, 5.10; N, 4.95.

(9b). ¹H NMR: δ 7.80 (s, 1H, NH); 7.36 (d, 1H, J_{H-6,H-5} = 8.2 Hz, H-6); 7.40-7.10 (m, 10 H, Ph); 5.97 (d, 1H, J_{H-1',H-2'} = 1.9 Hz, H-1'); 5.97 (dd, 1H, J_{H-5,NH} = 1.9 Hz, H-5); 4.80 (d, 1H, J_{gem}= 12.1 Hz, C<u>H2</u>Ph); 4.72 (d, 1H, J_{gem}= 12.1 Hz, C<u>H2</u>Ph); 4.59 (d, 1H, J_{gem}= 12.0 Hz, C<u>H2</u>Ph); 4.51 (d, 1H, J_{gem}= 12.0 Hz, C<u>H2</u>Ph); 4.32 (t, 1H, J_{H-3',H-4'} = J_{H-3',H-2'} = 3.0 Hz, H-3'); 4.04 (m, 3H, H-2', H-5', H-5''); 3.75 (ddd, 1H, J_{H-4',H-5'} = 10.1 Hz, J_{H-4',H-5''} = 6.1 Hz, H-4'). ¹³C NMR: δ 140.86 (C-6); 133.03-127.79 (Ph); 100.43 (C-5); 80.71 (C-1'); 77.68 (C-4'); 72.05 (CH2Ph); 71.72 (CH2Ph); 71.38 (C-3'); 65.88 (C-5'), 48.83 (C-2').

General procedure for reduction of the 2'-phenylselenenyl-pyranosyl nucleosides with tributyltin hydride. To a solution of 0.15 mmol of 2'-phenylselenenyl nucleoside in 2 mL of anhydrous benzene, 0.33 mmol (1 mL) of tri-butyltin hydride and 3 mg of 2.2'-azoisobutyronitrile (AIBN) were added at room temperature. Then the reaction was heated to reflux. When the starting material had disappeared (0.5-2 hours), the reaction mixture was cooled and evaporated dry. The resulting reaction crude was purified by flash chromatography.

1-(2'-Deoxy-3',4',6'-tri-O-acetyl-8-D-arabino-hexo-pyranosyl)uracil (11a). The reaction of compound 4a with tributyltin hydride was carried out following the general procedure. The reaction crude resulting was purified by flash chromatography (ethyl acetate-hexane 2:1) giving 46 mg (80%) of compound 11a.

¹³C NMR: δ 170.8 (CH₃<u>C</u>O); 170.1 (CH₃<u>C</u>O); 170.0 (CH₃<u>C</u>O); 163.3 (C-4); 150.0 (C-2); 139.1 (C-6); 103.3 (C-5); 79.0 (C-1'); 74.7 (C-4'); 70.1 (C-3'); 68.0 (C-5'); 61.9 (C-6'); 35.0 (C-2'); 20.5 (Me); 20.5 (Me); 20.4 (Me). Anal. Calcd. for C₁₆H₂₀N₂O₉; C, 50.03; H, 5.20; N, 7.29.

 $1-(2'-Deoxy-3',4',6'-tri-O-benzyl-B-D-arabino-hexo-pyranosyl)-uracil (11b) and <math>1-(2'-Deoxy-3',4',6'-tri-O-benzyl-\alpha-D-arabino-hexo-pyranosyl)-uracil (12).$ A mixture of compounds 4b and 5b was treated with tributylin hydride according with the general procedure. After 30 min. the reaction mixture was cooled and evaporated dry, obtaining 71 mg (90%) of a inseparable mixture of compounds 11b and 12. A small fraction of this mixture was purified by HPLC for identification purposes.

(11b). ¹³C NMR: δ 162.4 (C-4); 149.5 (C-2); 139.5 (C-6); 138.0-127.6 (Ph); 102.8 (C-5); 79.7 (C-1'); 79.6 (C-4'); 79.1 (C-3'); 77.0 (<u>C</u>H₂Ph); 75.5 (<u>C</u>H₂Ph); 73.4 (<u>C</u>H₂Ph); 71.9 (C-5'); 68.6 (C-6'); 36.0 (C-2').

(12). ¹³C NMR: δ 162.0 (C-4); 150.0 (C-2); 140.2 (C-6); 138.0-127.7 (Ph); 102.3 (C-5); 77.8 (C-1'); 73.4 (C-4'); 71.7 (C-3'); 73.4 (CH₂Ph); 71.2 (C-5'); 71.7 (CH₂Ph); 71.2 (CH₂Ph); 68.0 (C-6'); 30.3 (C-2').

1-(2'-deoxy-3',4',6'-tri-O-acetyl-B-D-lyxo-hexo-pyranosyl)-uracil (13a). Compound 7a was treated with tributyltin hydride following the general procedure. Flash chromatography (ethyl acetate-hexane 2:1) of the reaction crude gave 48 mg of 13a (83%)

Mp= 66-68 °C. $[α]_D^{23}$ +21.7° (c 0.6, CHCl₃). ¹³C NMR: δ 170.0 (CH₃<u>C</u>O); 169.9 (CH₃<u>C</u>O); 169.7 (CH₃<u>C</u>O); 162.7 (C-4); 149.7 (C-2); 139.1 (C-6); 103.3 (C-5); 79.4 (C-1'); 73.9 (C-4'); 68.0 (C-3'); 65.1 (C-5'); 61.8 (C-6'); 30.8 (C-2'); 20.7 (3xMe). Anal. Calcd. for C₁₆H₂₀N₂O₉: C, 50.03; H, 5.20; N, 7.29. Found: C, 49.71; H, 5.17; N, 7.08.

1-(2'-deoxy-3',4',6-tri-O-benzyl-B-D-lyxo-hexo-pyranosyl)uracil (13b). Compound 7b was treated with tributyltin hydride following the general procedure. Flash chromatography (ethyl acetate-hexane 1:1) of the reaction crude gave 60 mg of 13b (76 %).

Mp = 59-61 °C. $[α]_D^{20}$ +9.6° (c 0.5, CHCl₃). ¹³C NMR: δ 163.0 (C-4); 149.9 (C-2); 140.2 (C-6); 128.7-127.5 (Ph); 102.9 (C-5); 80.0 (C-1'); 76.7 (C-4'); 76.6 (C-3'); 74.6 (CH₂Ph); 73.6 (CH₂Ph); 71.8 (C-5'); 70.7 (CH₂Ph); 68.8 (C-6'); 31.8 (C-2'). Anal. Calcd. for C₃₁H₃₂N₂O₆: C, 70.45; H, 6.06; N, 5.30. Found: C, 69.42; H, 6.17; N, 5.52.

1-(2'-deoxy-3',4'-di-O-acetyl- α -D-erythro-pento-pyranosyl)uracil (14a). Compound 10a was treated with tributyltin hydride following the general procedure. Flash chromatography (ethyl acetate-hexane 2:1) of the reaction crude gave 42 mg of 14a (90%).

$$\begin{split} \mathbf{Mp} &= 70\text{-}72 \ ^{9}\text{C}. \ [\alpha]_{D}{}^{23} + 30.6^{\circ} \ (c \ \ 0.6, \ \text{CHCl}_{3}). \ ^{13}\text{C} \ \mathbf{NMR}: \ \delta \ 169.7 \ (3x\text{CH}_{3}\underline{O}); \ 162.9 \ (C-4); \ 149.8 \ (C-2); \\ 139.6 \ (C-6); \ 103.1 \ (C-5); \ 74.8 \ (C-1'); \ 73.4 \ (C-4'); \ 66.5 \ (C-3'); \ 66.3 \ (C-5'); \ 60.0 \ (C-6'); \ 32.9 \ (C-2'); \ 21.0 \\ (\text{Me}); \ 20.8 \ (\text{Me}); \ 20.6 \ (\text{Me}). \ \text{Anal. Calcd. for } C_{11}\text{H}_{16}\text{N}_{2}\text{O}_{7}: \ C, \ 50.00; \ \text{H}, \ 5.13; \ \text{N}, \ 8.97. \ \text{Found: } C, \ 48.84; \ \text{H}, \\ 5.09; \ \text{N}, \ 8.48. \end{split}$$

1-(2'-deoxy-3',4'-di-O-benzyl- α -D-erythro-pento-pyranosyl)uracil (14b). Compound 10b was treated with tributyltin hydride following the general procedure. Flash chromatography (ethyl acetate-hexane 1:1) of the reaction crude gave 52 mg of 14b (85%).

 $[\alpha]_D^{23}$ -36.6° (c, 6.5 CHCl₃). ¹³C NMR: δ 163.24 (C-4); 150.10 (C-2); 140.03 (C-6); 128.43-127.44 (Ph), 102.90 (C-5); 79.99 (C-1'); 75.19 (C-4'); 71.54 (<u>CH</u>₂Ph); 70.60 (<u>C</u>H₂Ph); 70.17 (C-3'); 66.72 (C-5'); 31.75 (C-2'). Anal. Calcd. for C₂₃H₂₄N₂O₅: C, 67.64; H, 5.88; N, 6.86. Found: C, 67.14; H, 5.76; N, 6.50.

Acknowledgement: This research was supported by a grant from DGICYT (Ministerio de Educación y Ciencia, Spain) Project PB89-0277

REFERENCES

- a) Mitsuya, H.; Yarchoan, R.; Broder, S. Science 1990, 249, 1533. b) "Design of anti-AIDS Drugs" DeClercq, E. Ed., Elsevier, New York, 1990. c) "AIDS: Modern Concepts and Therapeutic Challenges" Broder, S. Ed., Marcel Dekker, New York, 1987.
- a) Herdewijn, E.; Van Aerschot, A.; Balzarini, J.; De Clercq, E. Nucleosides & Nucleotides 1991, 10, 119. b) Van Aerschot, A.; Kerremans, L.; Balzarini, J.; De Clercq, E.; Herdewijn, E. Nucleosides & Nucleotides 1991, 10, 589. c) Bessodes, M.; Egron, M.J.; Filippi, J.; Antonakis, K. J. Chem. Soc., Perkins Trans. 1 1990, 3035.
- a) Périgaud, C.; Gosselin, G.; Imbach, J.L. J. Chem. Soc, Perkins Trans 1 1992, 1943. b) Baud, M.V.; Chauvis, C.; Lucas, M.; Imbach, J.L. Tetrahedron 1991, 47, 9993.
- a) Pitsch, S.; Wendeborn, S.; Jaun, B.; Eschenmoser, A. Helv. Chim. Acta 1993, 76, 2161, and
 references cited therein. b) Augustyns, K.; Rozenski, J.; Van Aerxchot, A.; Janssen, G.; Herdewijn, P. J. Org. Chem. 1993, 58, 2977.
- 5. Robins, M.J.; Wilson, J.S.; Hansske, F. J. Am. Chem. Soc. 1983, 105, 4059.
- a) Zorbach, W.W.; Munson, H.R.; Bhat, K.V.; J. Org. Chem. 1965, 30, 3955. b) Zorbach, W.W.; Munson, H.R. Synth. Proced. Nucleic Acid Chem. 1968, 1, 379. c) Lichtenthaler, F.W.; Kulikowski, T. J. Org. Chem. 1976, 41, 600.
- a) De las Heras, F.G.; Stud, M. Tetrahedron, 1977, 33, 1513. b) Kondo, T.; Nakai, H.; Goto, T. Tetrahedron 1973, 29, 1801. c) Leutzinger, E.E.; Meguro, T.; Townsend, L.B.; Shuman, D.A.; Schweizer, M.P.; Stewart, C.M.; Robins, R.K. J. Org. Chem. 1972, 37, 3695. d) Fuertes, M.; García Muñoz, G.; Madroñero, R.; Stud, M.; Rico. M. Tetrahedron 1972, 28, 623. e) Kondo, T.; Nakai, H.; Goto, T. Agr. Biol. Chem. 1971, 35, 1990. f) Ferrier, R.J. Ponpipom M.M. J. Chem. Soc. (C) 1971, 560. g) Leutzinger, E.E.; Robins, R.K.; Townsend, L.B. Tetrahedron Lett. 1970, 3751. h) Bowles, W.A.; Robins, R.K. J. Am. Chem. Soc. 1964, 86, 1256.
- 8. Bolitt, V.; Chaguir, B.; Sinou, D. Tetrahedron Lett. 1992, 33, 2481.
- a) Roush, W.R.; X-F. Liu Tetrahedron Lett. 1993, 34, 6829. b) Sedesta, D.P.; Roush, W.R. J. Org. Chem. 1992, 57, 4799. c) Kaila, N.; Grewal, G.; Franck, R.W. J. Org. Chem. 1992, 57 2084. d) Roush, W.R.; Lin, S.F. J. Org. Chem. 1991, 57, 5740. e) Ramesh, S.; Kaila, N.; Grewal, G.; Franck, R.W. J. Org. Chem. 1990, 55, 5. f) Preuss, R.; Schmidt, R.R. Synthesis 1988, 694.

- a) Beau, J.M.; Jaurand, G. Tetrahedron Lett. 1989, 30, 75. b) Kaye, A.; Neidle, S.; Reese, C.B. Tetrahedron Lett. 1988, 29, 2711. c) Jaurand, G.; Beau, J.M.; Sinaÿ, P. J. Chem. Soc. Chem. Commun. 1981, 527.
- 11. a) Thiem, J.; Klaffhe, W. J. Org. Chem. 1989, 54, 2006. b) Thiem, J.; Gerken, M. J. Org. Chem. 1985, 50, 954. c) Thiem, J.; Karl, H.; Schwenter, J. Synthesis 1978, 696.
- 12. Horton, D.; Priebe, W.; Sznaidman, M. Carbohydr. Res. 1990, 205, 71.
- a) Griffith, D.A.; Danishefsky, S.J. J. Am. Chem. Soc. 1990, 112, 5812. b) Friesen, R.W.; Danishefsky, S.J. Tetrahedron 1990, 46, 103.
- For 2'-deoxy-2'-phenylselenenyl furanosyl nucleosides see: Tong, W.; Xi, Z.; Gioeli, C.; Chattopadhyaya, J. *Tetrahedron*, 1991, 47, 3431
- 15. For conformational nomenclature rules in carbohydrate chemistry see: Schwarz, J.C.P. J. Chem. Soc. Chem. Commun. 1973, 505.
- 16. Rico, M.; Santoro, J. Org. Mag. Res. 1976, 8, 49.
- 17. For a detailed discussion of this effect see ref. 12
- 18. Kaluza, Z.; Pedersen, E.B.; Nielsen, C.M.; Chmieliwski, M. Acta. Chim. Sc., 1990, 44, 294.
- 19. Kiburis, J. J. Chem. Commun., Chem. Commun. 1975, 44.

(Received in UK 12 July 1994; revised 31 August 1994; accepted 2 September 1994)